101.425-06 – including *Taq* pol., IFU-01
101.425-06u – without *Taq* pol., IFU-02

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"Instructions for Use" (IFU)

Lot No.: 1F0 **Lot-specific information**

Olerup SSP® HLA-A*34

Product number:	**101.425-06 – including *Taq* pol.** **101.425-06u – without *Taq* pol.**
Lot number:	**1F0**
Expiry date:	**2019-10-01**
Number of tests:	**6**
Number of wells per test:	**11+1**
Storage - pre-aliquoted primers:	**dark at -20ºC**
- PCR Master Mix:	**-20ºC**
- Adhesive PCR seals	**RT**
- Product Insert	**RT**

This Product Description is only valid for Lot No. 1F0.

Complete product documentation consists of generic Instructions for Use (IFU), lot specific Product Insert, Worksheet and Certificate.

CHANGES COMPARED TO THE PREVIOUS *OLERUP* SSP® HLA-A*34 LOT (87Y)

The HLA-A*34 kit is updated for new alleles to enable separation of:

- Confirmed[1] alleles as listed in the IMGT/HLA database
- Polymorphisms in exons outside of the region encoding the peptide binding domain
- Null and Alternatively expressed alleles

The format of the Worksheet has been changed.

[1]As described in section Uniquely Identified Alleles.

The HLA-A*34 specificity and interpretation tables have been updated for the HLA-A alleles described since the previous *Olerup* SSP® HLA-A*34 lot was made **(Lot No. 87Y)**. The kit design is based on IMGT/HLA databased 3.26.0.

The primers of the wells detailed below have been exchanged, added or modified compared to the previous lot.

Well	5'-primer	3'-primer	rationale
8	-	-	Strength of control band has been optimized for increased yield.

Lot No.: 1F0 **Lot-specific information**

Well **12** contains Negative Control primer pairs, that will amplify more than 95% of the *Olerup* SSP® HLA Class I, DRB, DQB1, DPB1 and DQA1 amplicons as well as all the amplicons generated by the control primer pairs matching the human growth hormone gene.

HLA-specific PCR product sizes range from 75 to 200 base pairs.
The PCR product generated by the positive control primer pair is 430 base pairs.

Length of PCR product	**105**	**200**	**105**	**80**	**75**	**80**	**85**
5'-primer[1]	**164**	**340**	**440**	**45**	**45**	**43**	**36**
	5'-CAC3'	**5'-Agg3'**	**5'-TTA3'**	**5'-Tgg3'**	**5'-Tgg3'**	**5'-Tgg3'**	**5'-TAC3'**
							36
							5'-TAT3'
3'-primer[2]	**231**	**2nd I**	**507**	**59**	**58**	**57**	**47**
	5'-TgC3'	**5'-AAA3'**	**5'-TTg3'**	**5'-CTC3'**	**5'-ggC3'**	**5'-CTC3'**	**5'-ACA3'**
							48
							5'-gCA3'
							48
							5'-gCC3'
							52
							5'-TgT3'
A*	+	+	+				
B*	+	+	+				
C*	+	+	+				
DRB1				+	+		
DRB3				+	+		
DRB5				+			
DQB1					+		
DPB1						+	
DQA1							+

[1]The nucleotide position for HLA class I genes and the codon for HLA class II genes, in the 2nd or 3rd exon, matching the specificity-determining 3'-end of the primer is given. Nucleotide and codon numbering as on the www.ebi.ac.uk/imgt/hla web site. The sequence of the 3 terminal nucleotides of the primer is given.

[2]The nucleotide position for HLA class I genes and the codon for HLA class II genes, in the 2nd or 3rd exon or the 2nd intron, matching the specificity-determining 3'-end of the primer is given in the anti-sense direction. Nucleotide and codon numbering as on the www.ebi.ac.uk/imgt/hla web site. The sequence of the 3 terminal nucleotides of the primer is given.

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Lot No.: 1F0 **Lot-specific information**

PRODUCT DESCRIPTION

HLA-A*34 SSP subtyping

CONTENT

The primer set contains 5'- and 3'-primers for identifying the A*34:01 to A*34:17 alleles.

PLATE LAYOUT

Each test consists of 12 PCR reactions in a 16 well cut PCR plate. Wells 13 to 16 are empty.

1	2	3	4	5	6	7	8
9	10	11	NC	empty	empty	empty	empty

The 16 well cut PCR plate is marked with 'HLA-A*34' in silver/gray ink.
Well No. 1 is marked with the Lot No. '1F0'.
Wells 1 to 11 – HLA-A*34 high resolution primers.
Well 12 – Negative Control (NC).
A faint row of numbers is seen between wells 1 and 2 or wells 7 and 8 of the PCR trays. These stem from the manufacture of the trays, and should be disregarded.
The PCR plates are heat-sealed with a PCR-compatible foil.
Please note: When removing each 16 well PCR plate, make sure that the remaining plates stay sealed. Use a scalpel or a similar instrument to carefully cut the foil between the plates.

INTERPRETATION

Due to the sharing of sequence motifs between HLA-A alleles non-HLA-A*34 alleles will be amplified by some primer mixes. For further details see Specificity Table.

UNIQUELY IDENTIFIED ALLELES

All the HLA-A*34 alleles, i.e. **A*34:01 to A*34:17 alleles**, recognized by the HLA Nomenclature Committee in October 2016[1,2] will be amplified by the primers in the HLA-A*34 subtyping kit.

The HLA-A*34 kit enables separation of the confirmed HLA-A*34 alleles as listed in the IMGT/HLA database. An HLA allele is listed as confirmed by IMGT/HLA if it has been sequenced by more than a single laboratory or from multiple sources. Current allele confirmation status for HLA-A*34 alleles is listed below.

The HLA-A*34 kit also enables identification of polymorphisms in exons outside of the region encoding the peptide binding domain and of null and alternatively expressed alleles.

Lot No.: 1F0 **Lot-specific information**

[1]HLA-A alleles listed on the IMGT/HLA web page 2016-October-14, release 3.26.0, www.ebi.ac.uk/imgt/hla.

[2]Alleles that have been deleted from or renamed in the official WHO HLA Nomenclature up to and including the last IMGT/HLA database release can be retrieved from web page http://hla.alleles.org/alleles/deleted.html.

ALLELE CONFIRMATION STATUS

Allele	Status[1]	Allele	Status[1]	Allele	Status[1]
A*34:01:01	**Confirmed**	A*34:07	Unconfirmed	A*34:17	Unconfirmed
A*34:01:02	Unconfirmed	**A*34:08**	**Confirmed**		
A*34:02:01	**Confirmed**	A*34:09	Unconfirmed		
A*34:02:02	Unconfirmed	A*34:10N	Unconfirmed		
A*34:02:03	Unconfirmed	A*34:11	Unconfirmed		
A*34:02:04	Unconfirmed	A*34:12	Unconfirmed		
A*34:03	**Confirmed**	A*34:13	Unconfirmed		
A*34:04	Unconfirmed	A*34:14	Unconfirmed		
A*34:05	**Confirmed**	A*34:15	Unconfirmed		
A*34:06	Unconfirmed	A*34:16	Unconfirmed		

[1]Allele status "confirmed" or "unconfirmed" as listed on the IMGT/HLA web page 2016-October-14, release 3.26.0, www.ebi.ac.uk/imgt/hla.

RESOLUTION IN HOMO- AND HETEROZYGOTES

Results file with resolution in HLA-A*34 homo- and heterozygotes is available upon request.

Lot No.: 1F0

Lot-specific information

SPECIFICITY TABLE

HLA-A*34 SSP subtyping

Specificities and sizes of the PCR products of the 11+1 primer mixes used for HLA-A*34 SSP subtyping

Primer Mix	Size of spec. PCR product[1]	Size of control band[2]	Amplified HLA-A*34 alleles[3]	Other amplified HLA Class I alleles
1[4]	100 bp	**800 bp**	*34:01:01-34:06, 34:08, 34:10N-34:17	*01:13, 01:17, 01:176, 03:63, 03:88, 11:01:01:01-11:01:67, 11:01:69-11:11, 11:13-11:16, 11:20-11:27, 11:29-11:39, 11:41-11:52Q, 11:54-11:95, 11:97, 11:99N-11:105, 11:107-11:120, 11:122-11:158, 11:160-11:177, 11:179-11:246, 11:249, 11:251N, 25:02, 26:13, 26:19, 26:33, 29:66, 66:01:01:01-66:01:03, 66:04, 66:06-66:11, 66:13-66:14, 66:17-66:20, 66:22-66:24, 66:27N, 69:02, **C*07:335**
2[4]	110 bp	1070 bp	*34:01:01-34:01:02, 34:05, 34:11-34:12, 34:14, 34:16-34:17	*26:48, 26:69
3	195 bp	1070 bp	*34:02:01-34:02:04, 34:04, 34:07-34:10N, 34:13, 34:15	*03:248, 11:191
4[6]	135 bp	**800 bp**	*34:03, 34:06, 34:17	*03:01:19, 25:09, 26:14, 26:18, 26:28, 26:73, 26:112, 31:03-31:04, 66:22, 74:01:03
5	200 bp	**800 bp**	*34:04	*31:01:07, **B*15:82, B*15:260, B*40:186:01, C*03:186:01, C*12:57:02**
6	155 bp	1070 bp	*34:05	*02:91, 02:322, 03:94, **C*07:81, C*07:243, C*16:76**
7[7]	140 bp	1070 bp	*34:06	*26:18, 26:112[W], 31:03-31:04, 74:01:03
	215 bp		*34:07	*11:96
8[5]	200 bp	**800 bp**	*34:08	*01:51, 02:55, 02:644, 03:24, 25:03, 25:30, 26:20, 32:15, 68:71
	360 bp		*34:09	*02:135, 02:309, 02:454, 03:01:19, 25:13, 26:30, 26:65, 31:04, 66:02-66:03, 66:12, 66:16, 66:21, 66:25-66:26Q, 74:01:03
9	175 bp	1070 bp	*34:11	
10	185 bp	1070 bp	*34:10N	
11	175 bp	1070 bp	*34:01:01-34:09, 34:12-34:17	*02:309, 02:454, 03:01:19, 25:06, 26:09, 26:91, 31:03-31:04, 74:01:03
12[8,9]			**Negative Control**	

Lot No.: 1F0 **Lot-specific information**

[1]Alleles are assigned by the presence of specific PCR product(s). However, the sizes of the specific PCR products may be helpful in the interpretation of HLA-A*34 SSP typings.
When the primers in a primer mix can give rise to HLA-specific PCR products of more than one length this is indicated if the size difference is more than 20 base pairs. Size differences of 20 base pairs or less are not given. For high resolution SSP kits, the alleles listed are specified according to amplicon length.
Nonspecific amplifications, i.e. a ladder or a smear of bands, may sometimes be seen. GC-rich primers have a higher tendency of giving rise to nonspecific amplifications than other primers.
PCR fragments longer than the control bands may sometimes be observed. Such bands should be disregarded and do not influence the interpretation of the SSP typings.
PCR fragments migrating faster than the control bands, but slower than a 400 bp fragment may be seen in some gel read-outs. Such bands can be disregarded and do not influence the interpretation of the SSP typings.
Some primers may give rise to primer oligomer artifacts. Sometimes this phenomenon is an inherit feature of the primer pair(s) of a primer mix. More often it is due to other factors such as too low amount of DNA in the PCR reactions, taking too long time in setting up the PCR reactions, working at elevated room temperature or using thermal cyclers that are not pre-heated.

[2]The internal positive control primer pairs amplify segments of the human growth hormone gene. The internal positive control bands are 1070 or 800 base pairs respectively, well distribution as outlined in the table. Well number 1 contains the shorter, 800 bp, internal positive control band. The well distribution of the internal controls can help in orientation of the kit on gel photo, as well as allow for kit identification. In the presence of a specific amplification the intensity of the control band often decreases.

[3]For several HLA Class I alleles 1st and/or 4th exon(s) and beyond, as well as intron nucleotide sequences, are not available. In these instances it is not known whether some of the primers of the SSP sets are completely matched with the target sequences or not. Assumption is made that unknown sequences in these regions are conserved within allelic groups.

[4]HLA-specific PCR products shorter than 125 base pairs have a lower intensity and are less sharp than longer PCR products.

[5]In primer mix 8 the positive control band may be weaker than for other HLA-A*34 primer mixes.

[6]Primer mix 4 may have tendencies of unspecific amplification.

[7]Primer mix 7 may give rise to a long fragment of approx. 700 bp in some HLA-A alleles. This band should not be considered in the interpretation of HLA-A*34 typings.

[8]Primer mix 12 has a tendency to giving rise to primer oligomer formation.

[9]Primer mix 12 contains a negative control, which will amplify more than 95% of HLA amplicons as well as the amplicons generated by the control primer pairs matching the human growth hormone gene. HLA-specific PCR product sizes range from 75 to 200 base pairs and the PCR product generated by the HGH positive control primer pair is 430 base pairs.
'w', might be weakly amplified.

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Lot-specific information

PRIMER SPECIFICATION

Well No.	1	2	3	4	5	6	7	8	9	10	11
Length of spec.	100	110	195	135	200	155	140	200	175	185	175
PCR product							215	360			
Length of int.	800	1070	1070	800	800	1070	1070	800	1070	1070	1070
pos. control[1]											
5'-primer(s)[2]	282	270	363	423	78	445	103	102	423	415	423
	5' -CAg 3'	5' -AAA 3'	5' -ATA 3'	5' -gCT 3'	5' -TCC 3'	5' -TCT 3'	5' -CCT 3'	5' -ACA 3'	5' -gCT 3'	5' -ggT 3'	5' -gCT 3'
							423	341			
							5' -gCT 3'	5' -ggC 3'			
3'-primer(s)[3]	341	341	517	517	238	559	277	259	559	559	559
	5' -CgT 3'	5' -CgT 3'	5' -CgT 3'	5' -CgC 3'	5' -CCT 3'	5' -CgT 3'	5' -ggA 3'	5' -gTT 3'	5' -CgC 3'	5' -CgT 3'	5' -CgT 3'
							524	418			
							5' -CAC 3'	5' -gTC 3'			
Well No.	1	2	3	4	5	6	7	8	9	10	11

[1]The internal positive control primer pairs amplify segments of the human growth hormone gene. The internal positive control bands are 1070 or 800 base pairs respectively, well distribution as outlined in the table. Well number 1 contains the shorter, 800 bp, internal positive control band. The well distribution of the internal controls can help in orientation of the kit on gel photo, as well as allow for kit identification. In the presence of a specific amplification the intensity of the control band often decreases.

[2]The nucleotide position matching the specificity-determining 3'-end of the primer is given. Nucleotide numbering as on the www.ebi.ac.uk/imgt/hla web site. The sequence of the 3 terminal nucleotides of the primer is given.

[3]The nucleotide position matching the specificity-determining 3'-end of the primer is given in the anti-sense direction. Nucleotide numbering as on the www.ebi.ac.uk/imgt/hla web site. The sequence of the 3 terminal nucleotides of the primer is given.

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CELL LINE VALIDATION SHEET

HLA-A*34 SSP subtyping kit[2]

					Well										
					1	2	3	4	5	6	7	8	9	10	11
				Prod No.:	201557001	201557002	201557003	201557004	201557005	201557006	201557007	201778808	201557009	201557010	201557011
	IHWC cell line[1]		A*	A*											
1	9001	SA	*24:02		-	-	-	-	-	-	-	-	-	-	-
2	9280	LK707	*02:01		-	-	-	-	-	-	-	-	-	-	-
3	9011	E4181324	*01:01		-	-	-	-	-	-	-	-	-	-	-
4	9275	GU373	*30:01		-	-	-	-	-	-	-	-	-	-	-
5	9009	KAS011	*01:01		-	-	-	-	-	-	-	-	-	-	-
6	9353	SM	*02:01	*26:03	-	-	-	-	-	-	-	-	-	-	-
7	9020	QBL	*26:01		-	-	-	-	-	-	-	-	-	-	-
8	9025	DEU	*31:01		-	-	-	-	-	-	-	-	-	-	-
9	9026	YAR	*26:01		-	-	-	-	-	-	-	-	-	-	-
10	9107	LKT3	*24:02		-	-	-	-	-	-	-	-	-	-	-
11	9051	PITOUT	*29:02		-	-	-	-	-	-	-	-	-	-	-
12	9052	DBB	*02:01		-	-	-	-	-	-	-	-	-	-	-
13	9004	JESTHOM	*02:01		-	-	-	-	-	-	-	-	-	-	-
14	9071	OLGA	*31:01		-	-	-	-	-	-	-	-	-	-	-
15	9075	DKB	*24:02		-	-	-	-	-	-	-	-	-	-	-
16	9037	SWEIG007	*29:02		-	-	-	-	-	-	-	-	-	-	-
17	9282	CTM3953540	*03:01	*80:01	-	-	-	-	-	-	-	-	-	-	-
18	9257	32367	*33:03	*74:01	-	-	-	-	-	-	-	-	-	-	-
19	9038	BM16	*02:01		-	-	-	-	-	-	-	-	-	-	-
20	9059	SLE005	*02:01		-	-	-	-	-	-	-	-	-	-	-
21	9064	AMALA	*02:17		-	-	-	-	-	-	-	-	-	-	-
22	9056	KOSE	*02:01		-	-	-	-	-	-	-	-	-	-	-
23	9124	IHL	*02:01	*34:01	+	+	-	-	-	-	-	-	-	-	+
24	9035	JBUSH	*32:01		-	-	-	-	-	-	-	-	-	-	-
25	9049	IBW9	*33:01		-	-	-	-	-	-	-	-	-	-	-
26	9285	WT49	*02:05		-	-	-	-	-	-	-	-	-	-	-
27	9191	CH1007	*24:10	*29:01	-	-	-	-	-	-	-	-	-	-	-
28	9320	BEL5GB	*02:01	*29:02	-	-	-	-	-	-	-	-	-	-	-
29	9050	MOU	*29:02		-	-	-	-	-	-	-	-	-	-	-
30	9021	RSH	*30:01	*68:02	-	-	-	-	-	-	-	-	-	-	-
31	9019	DUCAF	*30:02		-	-	-	-	-	-	-	-	-	-	-
32	9297	HAG	*02:01		-	-	-	-	-	-	-	-	-	-	-
33	9098	MT14B	*31:01		-	-	-	-	-	-	-	-	-	-	-
34	9104	DHIF	*31:01		-	-	-	-	-	-	-	-	-	-	-
35	9302	SSTO	*32:01		-	-	-	-	-	-	-	-	-	-	-
36	9024	KT17	*02:06	*11:01	+	-	-	-	-	-	-	-	-	-	-
37	9065	HHKB	*03:01		-	-	-	-	-	-	-	-	-	-	-
38	9099	LZL	*02:17		-	-	-	-	-	-	-	-	-	-	-
39	9315	CML	*01:01	*03:01	-	-	-	-	-	-	-	-	-	-	-
40	9134	WHONP199	*02:07	*30:01	-	-	-	-	-	-	-	-	-	-	-
41	9055	H0301	*03:01		-	-	-	-	-	-	-	-	-	-	-
42	9066	TAB089	*02:07		-	-	-	-	-	-	-	-	-	-	-
43	9076	T7526	*02:06	*02:07	-	-	-	-	-	-	-	-	-	-	-
44	9057	TEM	*66:01		+	-	-	-	-	-	-	-	-	-	-
45	9239	SHJO	*23:01	*24:02	-	-	-	-	-	-	-	-	-	-	-
46	9013	SCHU	*03:01		-	-	-	-	-	-	-	-	-	-	-
47	9045	TUBO	*02:16	*03:01	-	-	-	-	-	-	-	-	-	-	-
48	9303	TER-ND	*02:01	*11:01	+	-	-	-	-	-	-	-	-	-	-

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[1]The provided cell line HLA specificities are retrieved from the http://www.ihwg.org/hla web site. The specificity of an individual cell line may thus be subject to change.

[2]The specificity of each primer solution in the kit has been tested against 48 well characterized cell line DNAs and where applicable, additional cell line DNAs.

No DNAs carrying the alleles to be amplified by primer solutions 4, 5, 7, 9 and 10 were available. The specificities of the primers in primer solutions 4 and 7 were tested by separately adding one additional 5'-primer, respectively three additional 3'-primers. In primer solutions 5 and 9 it was only possible to test the 5'-primers, the 3-primers were not possible to test. In primer solution 10 it was only possible to test the 3'-primer, the 5-primer was not possible to test.

In primer solution 7 one of the 3'-primers was not possible to test. One additional 5'-primer and one additional 3'-primer in primer mix 8 were tested by separately adding one additional 3'-primer or 5'-primer.

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ADDRESSES:

Manufacturer:
***Olerup* SSP AB**, Franzengatan 5, SE-112 51 Stockholm, Sweden.
Tel: +46-8-717 88 27
Fax: +46-8-717 88 18
E-mail: olerup-se@caredx.com
Web page: http://www.olerup.com

Distributed by:
Olerup GmbH, Löwengasse 47 / 6, AT-1030 Vienna, Austria.
Tel: +43-1-710 15 00
Fax: +43-1-710 15 00 10
E-mail: olerup-at@caredx.com
Web page: http://www.olerup.com

Olerup Inc., 901 S. Bolmar St., Suite R, West Chester, PA 19382
Tel: 1-877-OLERUP1
Fax: 610-344-7989
E-mail: olerup-us@caredx.com
Web page: http://www.olerup.com

For information on *Olerup* distributors worldwide, contact **Olerup GmbH**.